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INTRODUCTION

One of the challenges in the manufacturing of clinical-grade iPS cells is singlecell cloning, the step relevant to the standardization and automation of iPS cell culture protocols.

Here we report the results of our proof-of-concept study utilizing CellCelector™, high-throughput nanowell-based image-verified single cell cloning platform (HT-NIC), manufactured by ALS Automated Lab Solutions (Jena, Germany).



CellCELECTOR[™] PLATFORM

The CellCelector[™] system consists of automated single-cell picking instrument, image analysis software, and nanowell plates. Cell isolation and aspiration is realized by robotics system, for imaging automated inverted microscope is used.



NANOWELL MICROPLATES

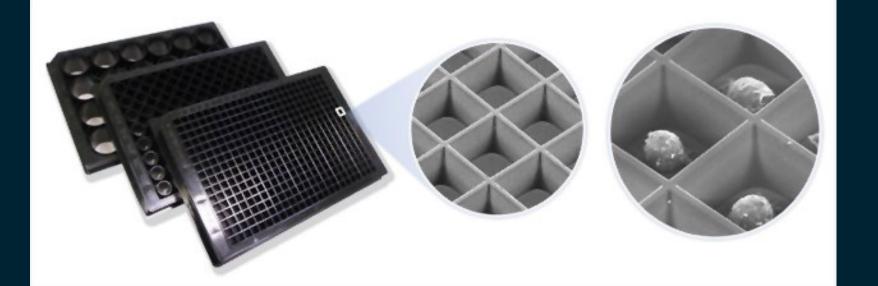
The nanowell plates facilitate isolation and identification of single cells, verification of monoclonality, assessment of the clones' outgrowth, and fast transfer of proliferating clones.

- diameter: 200 * 200 μm
- depth: 100 μm
- 132,000 nanowells per plate

ACKNOWLEDGEMENTS

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SINGLE-CELL CLONING OF INDUCED PLURIPOTENT STEM CELLS USING CELICELECTORTM INSTRUMENT AND NANOWELL PLATES

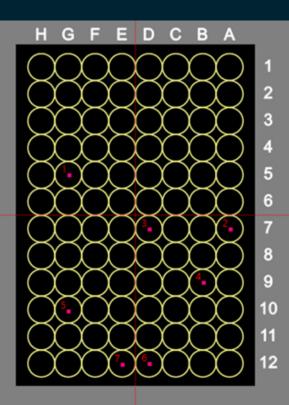
SINGLE CELL PICKING

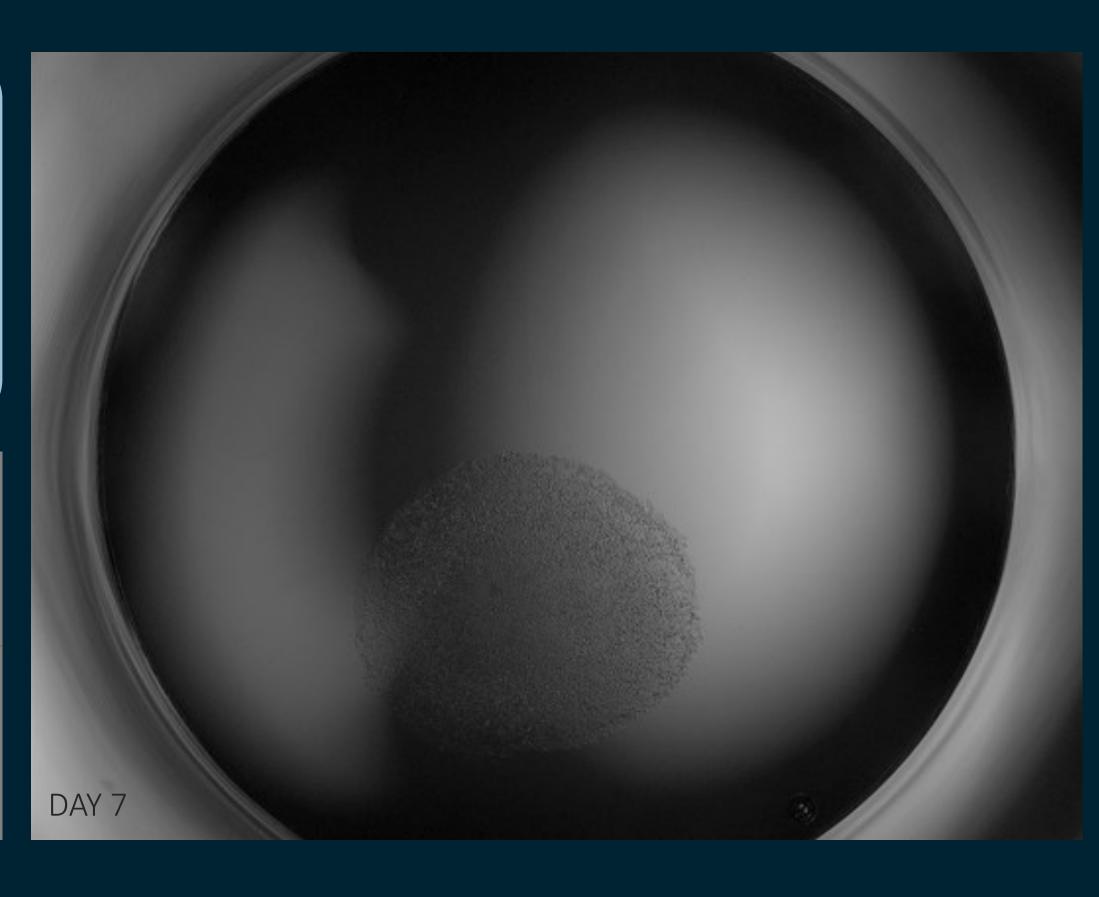
PROCEDURE

- iPS cells dissociation with Accutase[®] (Nacalai Tesque) and seeding on 24-well nanowell plate with density of 400 cells per well.
- Nanowell scanning with CellCelector[™] system and identification of nanowells containing single cells.
- Automated transfer of single cells to 96-well plate.
- Medium supplementation (cytoprotection) for 24 hours:
- Y-27632 (Fujifilm) [A1 H4];
- proprietary cocktail [A5 H8];
- CloneR (STEMCELL Technologies) [A9 H12].
- Culture at 37° C in humidified atmosphere with 5% CO₂ for 7 days.

RESULTS

- Microphotographs on culture day 7.
- Viable iPSC colonies:
- Y-27632 0 of 32 wells;
- cocktail 4 of 32 wells;
- CloneR 3 of 32 wells.

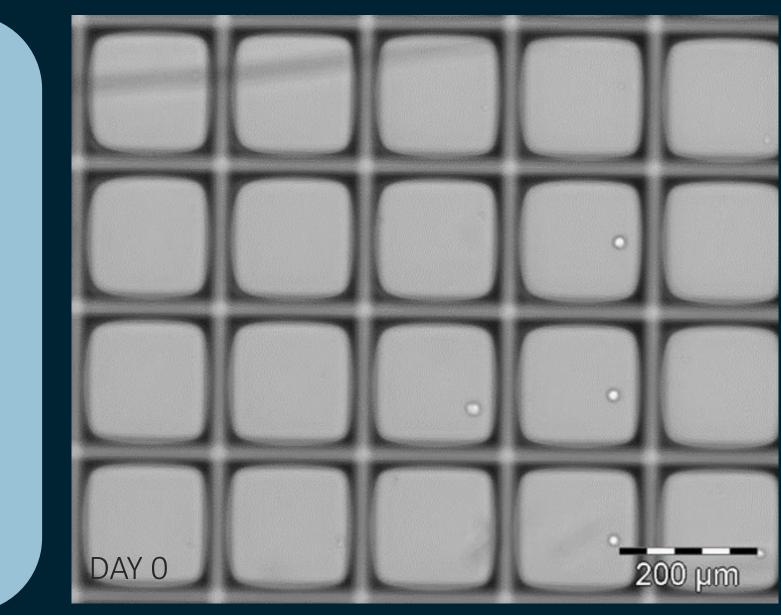




MATERIALS AND METHODS

CELLS

- iPS cells were generated from peripheral blood mononuclear cells (ePBMC, CTL) with Human iPS Cell Generation Episomal Vector Mix (TaKaRa) and 4d-Nucleofector[™] (Lonza).
- CULTURE SYSTEM
- Cells were cultured on iMatrix-511 substrate (Nippi) in StemFit AK03N medium (Ajinomoto). SINGLE CELL PICKING
- Single cells identified on nanowell plates were immediately transferred to 96-well plate (one cell per well). SINGLE CELL CLONING
- Seeded nanowell plates were scanned to identify nanowells containing single cells, the cells were cultured for 6 days, the plates scanned again, and suitable colonies transferred to 12-well plates.



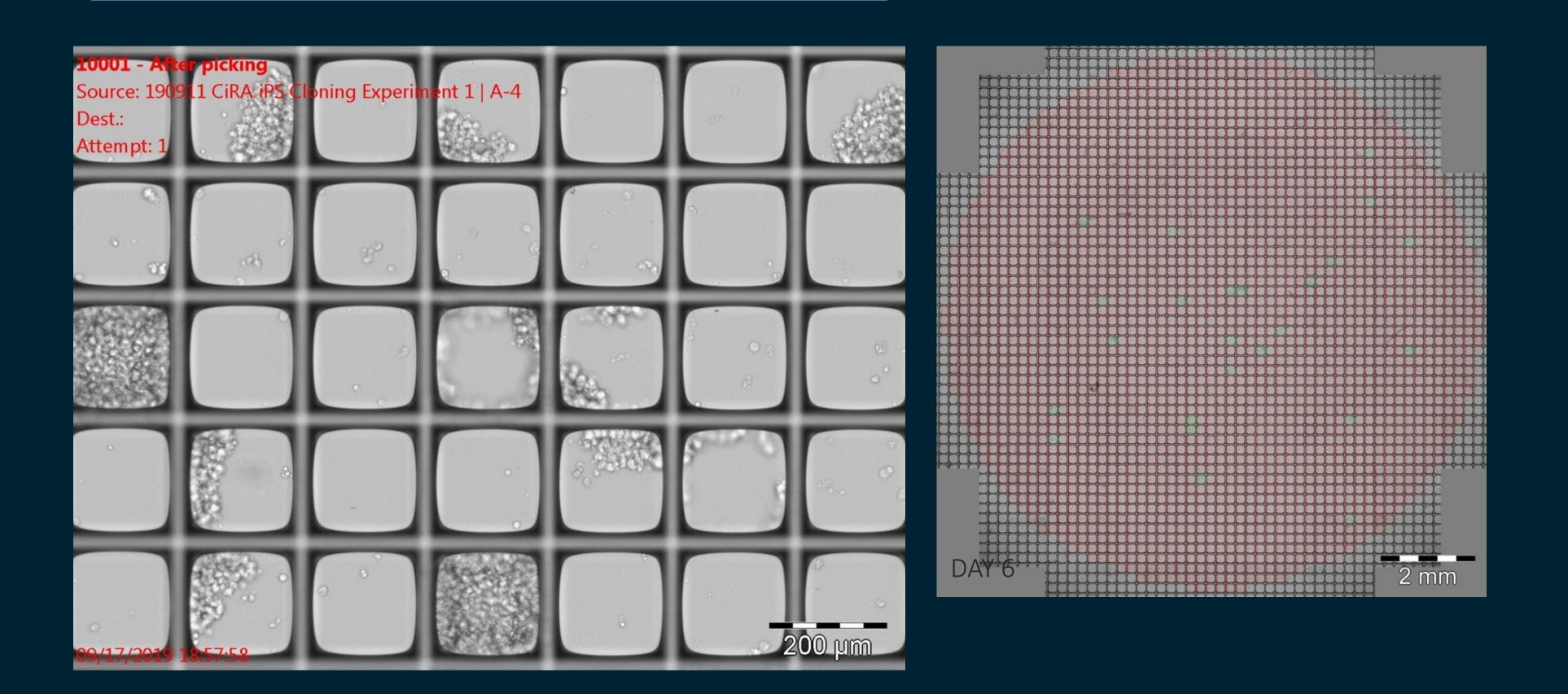
SINGLE CELL CLONING

PROCEDURE

- iMatrix-511 (Nippi) per well.
- Medium supplementation (cytoprotection) for 24 hours:
- Y-27632 (Fujifilm) [A2];
- proprietary cocktail [A4];
- CloneR (STEMCELL Technologies) [A6].
- Automated passage to 96-well plate:
- substrate iMatrix-511 (Nippi) 0.5 μg/cm²;
- dissociation Accutase[®] (Nacalai Tesque);
- colony picking with CellCelector[™] system.

RESULTS

to a 96-well plate.



CONCLUSIONS

- one standard culture plate.

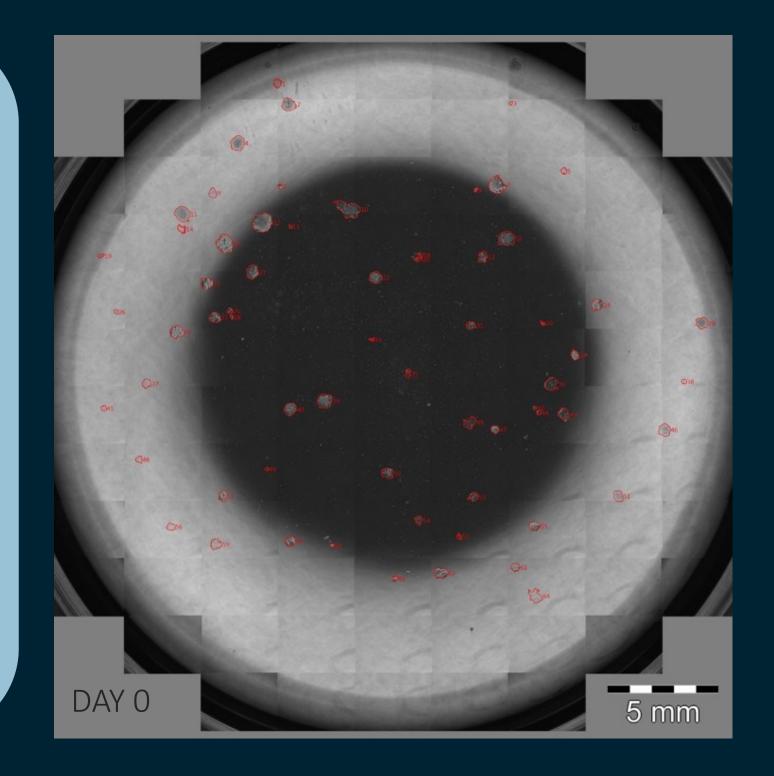
- characteristics for downstream applications.

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 iPS cells dissociation with Accutase[®] (Nacalai Tesque) and seeding on 24-well nanowell plate with density of 4,000 cells per well + 0.5 μ g of

• Culture at 37° C in humidified atmosphere with 5% CO₂ for 6 days.



Established iPSC colonies identified and successfully passaged

• CellCelector[™] system and nanowell microplates constitute excellent platform for generation of large number of iPS cell clones within

• High quality imaging system and image analysis software assures monoclonality of expanded iPS cells. Combination of CellCelector™ system and nanowell plates provides a tool for assessment of growth of iPS clones of desired